### AFRL-HE-WP-TP-2006-0021



### Air Force Research Laboratory

### Examination of HFE C282Y/H63D Heterozygotes as a Potential Human Modeling System for Low Level Liver Damage

Nathan A. Johnson Diane M. Todd Camilla A. Mauzy Julie A. Boyer Tracy A. Minnick Sean C. Stevens

Air Force Research Laboratory Human Effectiveness Directorate Applied Biotechnology Branch Wright-Patterson AFB, OH 45433-5707

K.A. Johnson 88 MDOS/SGOPC Wright-Patterson AFB OH 45433

March 2001

FINAL REPORT FOR THE PERIOD JUNE 1993 TO MARCH 2001

# 20060322008

Air Force Research Laboratory Human Effectiveness Directorate Biosciences and Protection Division Applied Biotechnology Branch Wright-Patterson AFB, OH 45433-5707

Approved for public release; distribution unlimited

### REPORT DOCUMENTATION PAGE

Form Approved OMB No. 0704-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Department of Defense, Washington Headquarters Services, Directorate for Information Operations and Reports (0704-0188), 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number. PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS.

1. REPORT DATE (DD-MM-YYYY) 2. REPORT TYPE		3. DATES COVERED (From - To)		
March 2001	Technical Paper	June 1993 to March 2001		
4. TITLE AND SUBTITLE		5a. CONTRACT NUMBER		
Examination of HFE C282Y/H6	N/A			
Human Modeling System for I	5b. GRANT NUMBER			
, <u>, , , , , , , , , , , , , , , , , , </u>	-	N/A		
		5c. PROGRAM ELEMENT NUMBER		
		62202F		
6. AUTHOR(S)		5d. PROJECT NUMBER		
Nathan H. Johnson, Diane M	. Todd, Camilla A. Mauzy, Julie A.	1710		
Boyer, Tracy A. Minnick		5e. TASK NUMBER		
		D4		
		5f. WORK UNIT NUMBER		
		1710D443		
7. PERFORMING ORGANIZATION NAME(S	S) AND ADDRESS(ES)	8. PERFORMING ORGANIZATION REPORT		
	, , , , , , , , , , , , , , , , , , , ,			
AND ADDRESS(ES)	,,	NUMBER		
AND ADDRESS(ES) Air Force Materiel Command		NUMBER		
AND ADDRESS(ES) Air Force Materiel Command Air Force Research Laboratory	Human Effectiveness Directorate	NUMBER		
AND ADDRESS(ES) Air Force Materiel Command Air Force Research Laboratory, Biosciences and Protection Div		NUMBER  AFRL-HE-WP-TR-2006-0021		
AND ADDRESS(ES) Air Force Materiel Command Air Force Research Laboratory	Human Effectiveness Directorate			
AND ADDRESS(ES) Air Force Materiel Command Air Force Research Laboratory, Biosciences and Protection Div	Human Effectiveness Directorate vision, Applied Biotechnology Branch,	AFRL-HE-WP-TR-2006-0021		
AND ADDRESS(ES) Air Force Materiel Command Air Force Research Laboratory, Biosciences and Protection Div Wright-Patterson AFB OH 45433	Human Effectiveness Directorate vision, Applied Biotechnology Branch,			
AND ADDRESS(ES) Air Force Materiel Command Air Force Research Laboratory, Biosciences and Protection Div Wright-Patterson AFB OH 45433	Human Effectiveness Directorate vision, Applied Biotechnology Branch,	AFRL-HE-WP-TR-2006-0021  10. SPONSOR/MONITOR'S ACRONYM(S)		
AND ADDRESS(ES) Air Force Materiel Command Air Force Research Laboratory, Biosciences and Protection Div Wright-Patterson AFB OH 45433	Human Effectiveness Directorate vision, Applied Biotechnology Branch,	AFRL-HE-WP-TR-2006-0021  10. SPONSOR/MONITOR'S ACRONYM(S)		
AND ADDRESS(ES) Air Force Materiel Command Air Force Research Laboratory, Biosciences and Protection Div Wright-Patterson AFB OH 45433	Human Effectiveness Directorate vision, Applied Biotechnology Branch,	AFRL-HE-WP-TR-2006-0021  10. SPONSOR/MONITOR'S ACRONYM(S)		
AND ADDRESS(ES) Air Force Materiel Command Air Force Research Laboratory, Biosciences and Protection Div Wright-Patterson AFB OH 45433	Human Effectiveness Directorate vision, Applied Biotechnology Branch,	AFRL-HE-WP-TR-2006-0021  10. SPONSOR/MONITOR'S ACRONYM(S) AFRL/HEPB		
AND ADDRESS(ES) Air Force Materiel Command Air Force Research Laboratory, Biosciences and Protection Div Wright-Patterson AFB OH 45433	Human Effectiveness Directorate vision, Applied Biotechnology Branch,	AFRL-HE-WP-TR-2006-0021  10. SPONSOR/MONITOR'S ACRONYM(S) AFRL/HEPB  11. SPONSOR/MONITOR'S REPORT		

Approved for public release; distribution is unlimited. Clearance AFRL/WS-06-0425, 15 Feb 06.

### 13. SUPPLEMENTARY NOTES

Poster Presentation at the Society of Toxicology, San Diego CA, March 2006

#### 14. ABSTRACT

Individuals heterozygous for hemochromatosis gene (HFE) mutations have been shown to demonstrate mild systemic iron-loading. In some studies, mildly elevated liver enzymes are noted. More sensitive testing methodologies are needed to assist in determining which heterozygous individuals may be at risk for future deleterious effects. Alpha Glutathione-S-Transferase (AGST) has been used to determine sub-clinical liver dysfunction and damage. However, AGST has not been used to examine the potential damage caused by mild iron loading. Whole blood/serum samples from individuals 20-50 years of age were collected from the Wright Patterson AFB clinical laboratory with only age and sex indicated. Genomic DNA was isolated from anticolagulated whole blood using the GFX Genomic Blood DNA Purification Kit (Amersham Biosciences). In two reactions, Exon 2 and Exon 4 of the HFE1 gene were amplified using primer sets HH63A/HH63B and HH1/HH5, respectively, in a standard PCR reaction (Accuprime PCR Kit, Invitrogen). The PCR products were digested either with MboI (Exon 2 containing the C186G mutation) or Rsal (Exon 4 containing the G845A mutation). The resultant restriction fragments were analyzed with matched controls on a 2% TBE agarose gel. Three groups of samples were identified (C282Y heterozygotes, H63D heterozygotes, and homozygous normal controls). Serum alanine aminotransferase, aspartate aminotransferase, gamma-glutamyl transferase, alkaline phosphatase, iron status, and AGST were analyzed on each sample. Samples from individuals possessing heterozygous C282Y mutation did not demonstrate a statistically significant elevation in AGST or other liver enzymes when compared to samples with no mutation. Similar results were found in individuals heterozygous for the H63D mutation. These results demonstrate that AGST activity may not be a good indicator for sub-clinical liver damage caused by increased loading of hepatic iron. Thus, the data does not support the use of human HFE C282Y/H63D heterzygtote samples in sub-clinical human liver dysfunction modeling.

#### 15. SUBJECT TERMS

Heterozygotes, liver damage, Alpha Glutathione-S-Transferase, Invitrogen, mutation, liver enzymes, hepatic iron

16. SECURITY CLASSIFICATION OF:		17. LIMITATION OF ABSTRACT	18. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON David R. Mattie	
a. REPORT U	b. ABSTRACT	c. THIS PAGE U		5	19b. TELEPHONE NUMBER (include area code) 937-904-9569

## EXAMINATION OF HFE C282Y/H63D HETEROZYGOTES AS A POTENTIAL HUMAN MODELING SYSTEM FOR LOW LEVEL LIVER DAMAGE

NH Johnson<sup>1</sup>, DM Todd<sup>1</sup>, CA Mauzy<sup>1</sup>, JA Boyer<sup>1</sup>, TA Minnick<sup>1</sup>, KA Johnson<sup>2</sup>, SC Stevens<sup>1</sup>

<sup>1</sup>Applied Biotechnology Branch-Human Effectiveness Directorate, Wright Patterson AFB, OH USA

<sup>2</sup>88 MDOS/SGOPC, Wright-Patterson AFB, OH, U.S.A.

### **Abstract**

Individuals heterozygous for hemochromatosis gene (HFE) mutations have been shown to demonstrate mild systemic iron-loading. In some studies, mildly elevated liver enzymes are noted. More sensitive testing methodologies are needed to assist in determining which heterozygous individuals may be at risk for future deleterious effects. Alpha Glutathione-S-Transferase (AGST) has been used to determine sub-clinical liver dysfunction and damage. However, AGST has not been used to examine the potential damage caused by mild iron loading. Whole blood/serum samples from individuals 20-50 years of age were collected from the Wright Patterson AFB clinical laboratory with only age and sex indicated. Genomic DNA was isolated from anticolagulated whole blood using the GFX Genomic Blood DNA Purification Kit (Amersham Biosciences). In two reactions, Exon 2 and Exon 4 of the HFE1 gene were amplified using primer sets HH63A/HH63B and HH1/HH5, respectively, in a standard PCR reaction (Accuprime PCR Kit, Invitrogen). The PCR products were digested either with MboI (Exon 2 containing the C186G mutation) or RsaI (Exon 4 containing the G845A mutation). The resultant restriction fragments were analyzed with matched controls on a 2% TBE agarose gel. Three groups of samples were identified (C282Y heterozygotes, H63D heterozygotes, and homozygous normal controls). Serum alanine aminotransferase, aspartate aminotransferase, gamma-glutamyl transferase, alkaline phosphatase, iron status, and AGST were analyzed on each sample. Samples from individuals possessing heterozygous C282Y mutation did not demonstrate a statistically significant elevation in AGST or other liver enzymes when compared to samples with no mutation. Similar results were found in individuals heterozygous for the H63D mutation. These results demonstrate that AGST activity may not be a good indicator for sub-clinical liver damage caused by increased loading of hepatic iron. Thus, the data does not support the use of human HFE C282Y/H63D heterzygtote samples in sub-clinical human liver dysfunction modeling.

### Introduction

Hereditary hemochromatosis (HHC) is an autosomal recessive disorder that results in increased iron absorption in affected individuals.(1) In 1996, the HFE gene was identified,(2) and mutations of this gene have been called the single most common gene disorder in Caucasian populations.(3) Multiple mutations of this gene have been identified and two major variants C282Y and H63D of the HFE gene have been

associated with clinical conditions.(1) Over time, increased iron absorption through the gastrointestinal tract in individuals affected with hemochromatosis (HHC) and this chronic iron loading state has been correlated with increased occurrence of a number of pathologic conditions including liver cancer and arthritis as well as lower states of well being with increase in fatigue. (4) The prevalence of C28Y homozygotes varies with ethnicity mixture of the population. Non-Hispanic Caucasians (0.44 percent) have the highest C28Y homozygotic prevalence(5). Although varying by population, the overall allele frequencies of the C282Y and H63D mutations have been reported penetrate at approximately 8 and 15 percent, respectively(6). Carriers of the C282Y mutations appear to have a stronger genetic correlation for prognosis dominated with ensuing pathology and age related clinical impact than H63D mutations.(1) Individuals heterozygous for either mutation normally have no clinical symptomology (1). Heterozygous individuals, especially those with a C282Y mutation, often have mild elevations of liver enzymes (i.e. alanine aminotransferase).(7) These mild elevations, along with evidence of synergistic relationships with dietary/environmental factors (i.e. clinical symptoms for C282Y mutation with alcohol intake), suggest that individuals heterozygous with HHC associated mutations may experience minimally intermittent or chronic mild liver damage.(7) Identification of individuals with mild HHC associated liver damage would provide a tremendous advantage to current treatments/diagnostic tools. The definitive diagnosis of liver associated damage attributed to HHC is biopsy.(1) An alternative is suggested due to the complications associated with biopsy. Alternatively, if standard iron levels (transferrin saturation and ferritin) are elevated, individuals may obtain therapeutic phlebotomies to lower iron burden.(1) These procedures, while certainly less invasive than biopsy, are cumbersome. Therefore, identification of the extent of the subpopulation exhibiting iron burden and liver damage at a "low level" or under normal dietary conditions is a worthy goal for screening this very large C282Y heterozygote population. There are many biomarkers currently available to study. One such biomarker is Alphaglutathione-S-transferase(AGST).(8) In the liver, AGST is enzyme located in the hepatocytes and has been shown in may acute and some chronic states to be elevated before traditional liver enzymes (alanine aminotransferase (ALT), aspartate aminotransferase (AST), gamma-glutamyl transferase (GGT), and alkaline phosphatase(AP)) have been elevated. (9-13) To date, AGST has not been used to investigate potential low level liver damage due to HFE mutations.

### Objective

The objective of this study was to evaluate serum AGST level as a possible indicator for low level human liver damage caused by mutations in the HFE gene.

### Materials and Methods

Whole blood/serum samples from individuals 20-50 years of age were collected from the Wright Patterson AFB clinical laboratory. Only age and sex were recorded. Genomic DNA was isolated from anticolagulated whole blood using the GFX Genomic Blood DNA Purification Kit (Amersham Biosciences). In two reactions, Exon 2 and Exon 4 of the HFE1 gene were amplified using primer sets HH63A/HH63B and HH1/HH5,

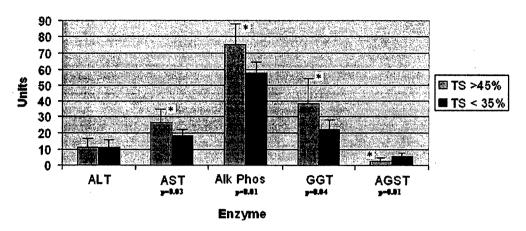
respectively, in a standard PCR reaction (Accuprime PCR Kit, Invitrogen). The PCR products were digested either with MboI (Exon 2 containing the C186G mutation) or RsaI (Exon 4 containing the G845A mutation). The resultant restriction fragments were analyzed with controls on a 2% TBE agarose gel. Three groups of samples were identified (C282Y heterozygotes, H63D heterozygotes, and homozygous normal controls). ALT, AST, GGT. AP, total iron and transferrin saturation (TS) were analyzed using standard procedures (Cobas Mira, Roche Diagnostics). AGST (Biotrin) was analyzed for each sample using microtiter plate antibody assay. Statistical analysis was performed using the Number Cruncher Statistical System. This study was approved by the 88th Medical Group IRB.

### Results

A contract which is a first or a second	Control (n=24)	C2B2y +/- (n=10)	H63D +/- (n=7)	Compound (n=6)
<b>基基的</b> 的L/的数据表	14.1	8.5	14.4	5.5
<b>CO</b> TTOYAGERS	26.3	19.8	21.7	16.5
AUSENTEVIOUS	72.35	59.5	58.6	51.6
SGI (1070)	38.2	19.3	20.6	20.6
eselective/ones	5.62	3.93	3.99	1.99
	175	113	155	129
HEKER BERRE	3B.3	35	47.7	47.8

	Age <39 (n=26)	Age >39 (n=26)	Male (n=14)	Female (n=39)
SISTE (IU/E) TARRES	11.2	12	17	9.5
2553 ((U/L) (1886)	20.4	24.7	24.3	22.1
BIREHTE (10)15	57	74	62	66
<b>CESURULO MEN</b>	20.5	37.2	29.7	28.4
ASSIMILITY OF	5.4	4.5	6.05	4.62
long(condit) (2.5	117	181	140	154
	36	41	41	38
AMELIAN BERTAIN	28	45	34	38

### Effect of TS% on Enzyme Activity



### Conclusion

Samples from individuals heterozygous for the C282Y mutation did not demonstrate a statistically significant elevation in AGST or other liver enzymes when compared to samples with no mutation. Instead, an average lower level value was determined in these individuals. No elevation was found in individuals heterozygous for the H63D mutation and in compound heterogygotes. These results demonstrate that elevated serum levels of AGST protein may not be a good indicator for sub-clinical liver damage caused by manifestations of mutations in the HFE gene. Surprisingly, individuals in this sampling of middle age individuals with mutations in the HFE gene did not have elevated iron status results. Examination of samples with elevated transferrin saturation percentage did show elevated liver enzyme activity with the exception being ALT and AGST, which showed a statistically significant decrease. These results do not support the initial hypothesis that AGST would be elevated in serum, could be used as detection of the condition, and could to demonstrate low level hepatic damage in individuals with HFE C282Y/H63D mutations. However, these data did show that lower levels of AGST were found in affected individuals. This counterintuitive finding may suggest a compensatory regulation of AGST or a lower hepatic AGST enzyme content in these individuals. Further investigation is merited, since increases in AGST levels are presently assumed as an event (an acute) hepatic event, where this appears not to be the clear case for chronic liver iron handling/stressor conditions.

### Reference List

1. Hanson EH, Imperatore G, Burke W. HFE gene and hereditary hemochromatosis: a HuGE review. Human Genome Epidemiology. Am J Epidemiol 2001;154:193-206.

- 2. Feder JN, Gnirke A, Thomas W, Tsuchihashi Z, Ruddy DA, Basava A. The discovery of the new haemochromatosis gene. 1996. J Hepatol 2003;38:704-9.
- 3. Fuchs J, Podda M, Packer L, Kaufmann R. Morbidity risk in HFE associated hereditary hemochromatosis C282Y heterozygotes. Toxicology 2002;180:169-81.
- 4. McDonnell SM, Preston BL, Jewell SA, Barton JC, Edwards CQ, Adams PC, Yip R. A survey of 2,851 patients with hemochromatosis: symptoms and response to treatment. Am J Med 1999;106:619-24.
- 5. Adams PC, Reboussin DM, Barton JC, McLaren CE, Eckfeldt JH, McLaren GD et al. Hemochromatosis and iron-overload screening in a racially diverse population. N Engl J Med 2005;352:1769-78.
- 6. Jackson HA, Carter K, Darke C, Guttridge MG, Ravine D, Hutton RD et al. HFE mutations, iron deficiency and overload in 10,500 blood donors. Br J Haematol 2001;114:474-84.
- 7. Powell LGKMSKK. Diagnosis of Hemochromatosis. Annals of Internal Med 1998;129:925-31.
- 8. Rees GW, Trull AK, Doyle S. Evaluation of an enzyme-immunometric assay for serum alpha-glutathione S-transferase. Ann Clin Biochem 1995;32 (Pt 6):575-83.
- 9. Trull AK, Facey SP, Rees GW, Wight DG, Noble-Jamieson G, Joughin C et al. Serum alpha-glutathione S-transferase--a sensitive marker of hepatocellular damage associated with acute liver allograft rejection. Transplantation 1994;58:1345-51.
- 10. Clarke H, Egan DA, Heffernan M, Doyle S, Byrne C, Kilty C, Ryan MP. Alphaglutathione s-transferase (alpha-GST) release, an early indicator of carbon tetrachloride hepatotoxicity in the rat. Hum Exp Toxicol 1997;16:154-7.
- 11. Hughes VF, Trull AK, Gimson A, Friend PJ, Jamieson N, Duncan A et al. Randomized trial to evaluate the clinical benefits of serum alpha-glutathione S-transferase concentration monitoring after liver transplantation. Transplantation 1997;64:1446-52.
- 12. Platz KP, Mueller AR, Muller C, Wenig M, Schumacher G, Steinmuller T et al. Indications for determination of alpha-glutathione-s-transferase after liver transplantation. Transplant Proc 1999;31:462-4.
- 13. Platz KP, Mueller AR, Haller GW, Muller C, Wenig M, Neuhaus R et al. Determination of alpha- and Pi-glutathione-S-transferase will improve monitoring after liver transplantation. Transplant Proc 1997;29:2827-9.